

Schneider-Maunoury L, Lefebvre V, Ewers RM, Medina-Rangel GF, Peres CA, Somarriba E, Urbina-Cardona N, Pfeifer M. [Abundance signals of amphibians and reptiles indicate strong edge effects in Neotropical fragmented forest landscapes](#). *Biological Conservation* 2016, 200, 207-215.

Copyright:

© 2016. This manuscript version is made available under the [CC-BY-NC-ND 4.0 license](#)

DOI link to article:

<http://dx.doi.org/10.1016/j.biocon.2016.06.011>

Date deposited:

29/06/2016

Embargo release date:

25 December 2017



This work is licensed under a [Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International licence](#)

1 Abundance signals of amphibians and reptiles indicate strong edge effects in Neotropical
2 fragmented forest landscapes

3 Laure Schneider-Maunoury^{1,2} (laure.schneider-maunoury@ens.fr)

4 Veronique Lefebvre¹ (v.lefebvre@imperial.ac.uk)

5 Robert M. Ewers¹ (r.ewers@imperial.ac.uk)

6 Guido. F. Medina-Rangel³ (gmedinar@unal.edu.co)

7 Carlos A. Peres⁴ (c.peres@uea.ac.uk)

8 Eduardo Somarriba⁵ (esomarri@catie.ac.cr)

9 Nicolás Urbina-Cardona⁶ (urbina-j@javeriana.edu.co)

10 Marion Pfeifer⁷ (marion.pfeifer@newcastle.ac.uk)

11

12 ¹ Imperial College, Silwood Park Campus, Buckhurst Road, Ascot SL57PY, UK

13 ² École Normale Supérieure de Paris, 45 rue d'Ulm, 75005 Paris, France (present address)

14 ³ Grupo de Biodiversidad y Conservación, Reptiles, Instituto de Ciencias Naturales, Universidad

15 Nacional de Colombia, Ciudad Universitaria, Edificio 425, Bogotá D. C., Colombia

16 ⁴ School of Environmental Sciences, University of East Anglia, Norwich NR47TJ, UK

17 ⁵ Programa de Agroforestería y Agricultura Sostenible, Sede Central, Centro Agronomico Tropical

18 de Investigacion y Ensenanza, 7170 Cartago, Turrialba 30501, Costa Rica

19 ⁶ Ecology and Territory Department, School of Rural and Environmental Studies, Pontificia

20 Universidad Javeriana, Bogotá D. C., Colombia

21 ⁷ School of Biology, Newcastle University, Newcastle NE17RU, UK

22

23 Corresponding author: Laure Schneider-Maunoury (+33 6 74 35 94 99)

24

25

26

27 **Abstract**

28 Fragmentation and habitat loss contribute considerably to global declines of amphibians and
29 reptiles. However, few studies focus on forest edges, created during the fragmentation process, as
30 proximate drivers of the local demographic structure of populations. Here, we use abundance data
31 of amphibians and reptiles to study their responses to forest edges in nine fragmented forested
32 landscapes of the Neotropics. Species-specific abundance data were collected in plots established at
33 varying distances from their respective nearest forest edge. We tested for edge effects on the
34 abundance of species, and used curve clustering techniques to group species with similar edge
35 responses, i.e. species with either increasing or decreasing abundance from the matrix towards the
36 forest interior. We also grouped species that showed no change in abundance with respect to the
37 nearest forest edge and those whose abundance response was unimodal, peaking in either forest
38 habitat or the surrounding matrix habitat. We found that 96% of all amphibians and 90% of all
39 reptiles showed an edge response, with the abundance of 74.5% of amphibians and 57.3% of
40 reptiles decreasing with increasing proximity to forest edges. However, species-specific edge
41 effects were not always consistent, with some species having opposite edge responses when
42 measured in different landscapes. The depth of edge effects exhibited by forest species, i.e. species
43 that increased in abundance in the forest interior, extended up to one kilometre away from forest
44 edges. We show that the median edge effect on forest species extends to 250m within the forest
45 interior, indicating that tropical forest patches with a mean diameter $< 500\text{m}$ (minimum area \approx
46 78ha) are unsuitable for half of forest-dependent species considered in this study.

47

48 **Keywords:** forest fragmentation, edge influence, species abundance, tropical forest, herpetofauna

49

50 **1 Introduction**

51 Forest fragmentation acts on top of forest loss, as continuous forest habitats are broken up into
52 increasingly smaller forest patches that are becoming increasingly isolated from each other (Fahrig,

2003). Fragmentation ultimately results in edge-dominated forest fragments, thereby edges or boundary zones differ structurally and functionally from both the original forest and the surrounding non-forest matrix (Saunders, 1991; Murcia, 1995; Cadenasso et al., 1997; Harper et al., 2005). Boundary zones are characterised by abiotic gradients such as wind, temperature or radiation and water fluxes (Didham, 2010), which cause first-order biological changes including altered species composition, distribution and abundance (Holt and Keitt, 2005; Ewers and Didham, 2006a). These biotic effects result in second-order biological changes, such as changes in trophic interactions (Fagan et al., 1999; Holt and Keitt, 2005) and parasitism (Schlaepfer and Gavin, 2001). Both abiotic gradients and biotic changes are examples of ecological phenomena grouped together under the broad concept of “edge effects”.

Edge effects can be quantified by their magnitude, i.e. the difference between the lowest and the highest value of a variable measured across a forest edge, and by their extent, i.e. the distance over which the change can be detected (Ewers and Didham, 2006b). Both measures may vary with the quality of the matrix and forest habitat, the latter depending for example on structure and age of the forest patch, shaping the spatially variable patch-matrix contrast (Laurance et al., 2011).

Species respond to the edge differently depending on life history traits, including those linked to habitat specialisation, body size and dispersal capacity (Saunders et al., 1991; Ewers and Didham, 2006a, Stork et al., 2009). Specialisation allows species to adapt to micro-environments as specialized species have a narrower niche breadth, but it can exacerbate the effect of habitat loss (Houlahan and Findlay, 2003; Williams et al., 2008). Indeed, it may render species more vulnerable to habitat changes because forest fragmentation can reduce the probability that the species and its niche are both maintained in the remaining forest fragments (Harris and Silva-Lopez, 1992; Henle et al., 2004). Larger-sized animals are reportedly been more sensitive to fragmentation (Crooks, 2002; Henle et al., 2004; Pineda and Halfpeter, 2004), although empirical studies report a range of responses (e.g. Suazo-Ortuno et al., 2008; Mendenhall et al., 2014). Dispersal capacity affects the

79 species ability to travel through suboptimal habitats, less able to provide resources or protection
80 from predators, e.g. low tree cover matrix habitat for forest species (McGarigal and Cushman,
81 2002).

82 Biodiversity responses to anthropogenic land use changes are often analysed as species
83 extinctions following habitat loss (Wearn et al., 2012). However, such responses are only the last
84 step of a decline in abundance. Abundance signals in response to land use change, and in particular
85 fragmentation, can be detected earlier, prior to actual species loss, and can be used to identify
86 vulnerable species that are at risk from local extinctions. This information can then be used to
87 design and implement conservation and management actions aimed at reversing local abundance
88 declines. However, modelling the response of abundance-based biodiversity indicators to local
89 drivers (e.g. fragmentation) requires high quality input data (Bellard et al. 2012) and would benefit
90 from a more mechanistic understanding of driver-response relationships at landscape scale.

91 The abundance and species diversity and richness of mammals and birds typically decrease
92 with increasing fragmentation (Andren, 1994; Laurance et al., 2011), even if some species are
93 disturbance-adapted and can be found in the matrix (e.g. Pardini, 2004). Amphibians and reptiles
94 are considered key indicators of environmental changes (Schlaepfer and Gavin, 2001; Blaustein
95 and Bancroft, 2007), with habitat loss and degradation known to contribute to their observed
96 worldwide declines (Gibbons et al., 2000; Houlahan and Findlay, 2003; Stuart et al., 2004;
97 Cushman, 2006). Yet, comparatively little is known about their responses to fragmentation,
98 especially when edge effects are considered (McGarigal and Cushman, 2002; Gardner et al., 2007a;
99 Carvajal-Cogollo and Urbina-Cardona, 2008, 2015; Suazo-Ortuño et al., 2008). Edges are typically
100 characterised by more open canopies, leading to reduced moisture and increased maximum daily
101 temperatures (Didham and Lawton, 1999; Hardwick et al., 2015). These altered microclimates are
102 particularly likely to affect abundance and distribution of amphibians (Lehtinen et al., 2003), as
103 they respire primarily through their moist skin and may thus be more sensitive to desiccation in drier
104 environments. Low dispersal ability of both amphibians (Gibbs, 1998; Demaynadier and Hunter,

105 1999; Cushman, 2006) and reptiles (Araujo and Pearson, 2005) is likely to further amplify such
106 edge effects.

107 Lehtinen et al. (2003) used randomisation techniques on species presence-absence data
108 across forest fragments in Madagascar to identify edge responses in amphibians and reptiles,
109 showing that they can display edge and non-edge avoiding strategies. However, their approach
110 could not account for spatial variation in abundance, which may be a more sensitive to
111 fragmentation and hence a more reliable indicator of edge effects on herpetofauna. This is
112 important, as species may show a gradient in their sensitivity to the edge effect, reflecting their
113 varying ability to exploit unique combinations of ecological conditions (Schlaepfer and Gavin,
114 2002). For example, species declining in abundance from the forest interior to the forest edge may
115 be mislabelled as non-sensitive to fragmentation based on their presence-absence pattern, even
116 though their abundance suggests the likelihood of a local extinction with continuing fragmentation
117 and loss of forest core habitat.

118 Here, we analyse the abundance responses of 43 amphibian and 61 reptile species to the
119 nearest forest edge in fragmented, human-modified Neotropical landscapes like plantations,
120 primary and secondary forests. We hypothesize that abundance of forest and non-forest species
121 show consistent signals in response to the forest edge, indicating causal impacts of habitat
122 fragmentation, and that we can exploit these signals to monitor the ecological integrity of forests in
123 the landscape. We quantify how far edge effects extend to within forest and matrix habitat by
124 computing the depth of edge influence; and we then use this depth of edge influence to test to what
125 extent habitat suitability decreases for forest species due to fragmentation. Finally, we compare
126 edge responses of species across multiple landscapes to investigate whether edge responses are
127 species-specific or whether they vary among fragmented landscapes, which would indicate that
128 landscape specific characteristics can modify the edge effect.

129

130

2 Material and Methods

2.1 *Species abundance*

We extracted amphibian and reptile datasets obtained at 11 Neotropical landscapes from the BIOFRAG database (Pfeifer et al., 2014) in February 2014, including data from both published and unpublished sources. We subsequently excluded two datasets from our analysis, one of which because of the small number of plots ($N = 9$) and another one because it was conducted in urban environment. The datasets contain the raw abundance data, i.e. the number of individuals of each species found in each given plot along survey seasons. One dataset solely focussed on measuring abundance of reptiles (Table 1). We concentrated on amphibians and reptiles identified at the level of species (85.7 % of all observations, 249 species retained). We excluded rare species from our analyses, defined as species whose abundance did not exceed three individuals at any plot, as their abundance patterns could not be analysed statistically. Therefore, we retained a total of 104 species (43 amphibian and 61 reptile species) for all subsequent analyses (see Supplementary material 1).

2.2 *Study landscapes*

The retained datasets originate from sampling in nine fragmented forest landscapes from Central and South America, whose characteristics are summarized in Table 1. The datasets encompass a variety of matrix types, including pasture, secondary regrowth or plantations. The sampling effort varies across datasets, with the number of sampling plots ranging from 15 to more than 150, and the number of known species sampled in these plots from 3 to 40.

2.3 *Delineating the forest edge*

Location of the forest edge was estimated from high spatial resolution tree cover maps generated from Landsat data between 2000 and 2003 (Hansen et al., 2013; Fig. 1A). We classified the continuous tree cover values as “forest” and “non-forest” using a three-step procedure: (1) homogeneous zones (tree cover variation $< 5\%$) were delineated using morphological segmentation

(marker controlled watershed algorithm, Meyer and Beucher, 1990; Fig. 1B), (2) maps were transformed so that pixels in each homogeneous zones were given the value of the average tree cover in the zone and (3) we computed the value of the tree cover threshold between “forest” and “non-forest” classes so that the intra-class variance of the darkest (high tree cover) and brightest (low tree cover) regions of the image was minimized (Otsu’s threshold, Otsu, 1979; Fig. 1C). Tree cover values above threshold were classified as “forest” and tree cover values below threshold as “non forest”. The delineation steps prior to thresholding ensure that neighbouring pixels of similar tree cover (differing by less than 5%) are classified within the same category. Therefore, the location of the forest edge, i.e. the interface between “forest” and “non forest” zones, always corresponds to an edge in the landscape.

As the temporal difference between tree-cover map acquisition and species abundance data sampling may introduce an uncertainty in our analysis, we examined whether the studied landscapes had not been significantly altered during this lag period for different studies (see Supplementary Material 2).

2.4 Estimation of edge influence

We used the distance to nearest edge as a proxy measure for edge influence. The distance transform for each forest/non-forest map was computed using the Matlab “bwdist” function implemented from Maurer et al. (2003). The distance transform map contains for each forest pixel its Euclidean distance to the nearest non-forest pixel (as a positive value), and for each non-forest pixel its Euclidean distance to the nearest forest pixel (as a negative value). We then extracted the distance to the nearest edge for each measurement plot from the distance transform map.

2.5 Computing species’ response curves to edges

The measured abundance of each species was plotted as a function of distance to the nearest edge (hereafter referred to as “edge response curve”). We searched for shape similarity and natural

grouping within edge response curves using curve clustering. A curve clustering algorithm regroups functions of the same domain (here distance to edge) by shape and location. The implementation we used (Gaffney, 2004) requires a function model to characterise and optimise each cluster. Equation 1 gives the function model we used to cluster edge response curves: a sum of an increasing and a decreasing logistic function. We developed this model to best represent prior knowledge on edge response curves (Ewers and Didham, 2006b): response curves have a sigmoidal shape (species abundance reaches a horizontal asymptote where the edge influence is negligible) and response curves may be monotonic or go through a maximum.

$$\eta = \frac{J}{1 + e^{-B(x-M)}} + \frac{-J}{1 + e^{-B(x-(M+addM))}}$$

Equation 1

Equation 1 is a sum of two sigmoids (logistic functions), modelling the species abundance η as a function of the distance to the nearest forest edge x . The amplitude J is constrained between 0 and maximum abundance, so that the first sigmoid increases and the second sigmoid decreases. B is the steepness of the curve, termed “growth rate” or “de-growth rate” for increasing or decreasing sigmoids respectively. The value of B is positive and is constrained to be below 0.1 to model relatively slow transitions (Ewers and Didham, 2006b). M is the distance to edge for which the growth rate of the first sigmoid is maximum (i.e. M is the location of the inflection point of the sigmoid curve, where the first derivative reaches a maximum and the second derivative goes through zero). We refer to the parameter M as the “growth distance”. M is constrained to be within three times the observed distance range: $\text{min distance} - (\text{max distance} - \text{min distance}) < M < \text{max distance} + (\text{max distance} - \text{min distance})$. $M+addM$ is the inflection point of the second (decreasing) sigmoid, termed “de-growth distance”. $addM$ is positive and lower than $3 \times (\text{max distance} - \text{min distance})$ so that de-growth follows growth and species abundance goes through a

207 maximum (and never a minimum).

208 Because the distance to the nearest edge x varies between the minimum and maximum of
209 measured distances, this model (Equation 1 with constrained parameters) can return either an
210 increasing sigmoid (with $M + \text{add}M \gg \text{max distance}$, Fig. 2 responses 3 and 4), a decreasing
211 sigmoid ($M \ll \text{min distance}$, Fig. 2 responses 1 and 2), a peaking curve (with M and $M + \text{add}M$
212 within the observed distance range, Fig. 2 responses 5 to 7), or an almost flat curve (M and
213 $M + \text{add}M$ outside, Fig. 2 response 0).

214

215 *2.6 Summarising and comparing edge response curves*

216 We analysed patterns in species edge response curves across datasets by classifying the clusters
217 obtained in the previous step into 8 pre-defined possible response types (Fig. 2). The parameters of
218 the initial clusters were chosen to obtain a set of decreasing, increasing, peaking and flat curves
219 regularly spaced over the range of measured distance. From preliminary testing we estimated that 8
220 clusters were sufficient to represent the variability of the data. These 8 response types were further
221 re-grouped into three broader categories (Fig. 2) based on the optimised parameters of each cluster
222 (i.e. whether growth and de-growth distances were positive, negative or out of the distance range):
223 species showing no response to the edge (response 0 or “no response”, i.e. generalist species),
224 species decreasing in abundance from the matrix to the forest interior (responses 1 and 2 or “non-
225 forest species”), and species increasing in abundance from the matrix to within the forest interior
226 (responses 3, 4 and 5 or “forest species”). The remaining two possible response types (6 and 7)
227 were never observed.

228 In a second step, we quantified the impact of the edge on the abundance of the species
229 (termed “depth of edge influence”) as the location of the minimum of the second derivative of the
230 response curve, or the location of the closest minimum to the edge when there are two minima
231 (Ewers and Didham, 2006b). The location of the minimum of the second derivative corresponds to
232 the distance for which the influence of the edge on the species abundance becomes negligible (cut-

off point). Species present in several datasets were treated as distinct species. This allows us to test whether their edge response curve depends on the landscape in which they are measured.

3 Results

Among the 104 species analysed, six amphibian and 11 reptile species were present in more than one landscape, resulting in a total of 133 observed response curves (51 for amphibians and 82 for reptiles). A significant relationship with distance to the nearest edge was observed for 92.5% of the curves, with only two of the 51 amphibian curves and eight of the 82 reptile curves failing to show any pattern in their abundance responses (Fig. 3).

3.1 Grouping species according to their edge response curves

Response curves of species analysed matched six of eight possible clusters of edge response types (Fig. 2). In total, 85 species showed response curves that resembled those of forest species (38 amphibian and 47 reptile responses), whilst response curves of 38 species (11 amphibian and 27 reptile responses) resembled those of non-forest species. Edge response type 4, i.e. forest species that increase in abundance from the forest edge to forest interior and are absent from the matrix, was the most common edge response (Fig. 3).

The mean depth of edge influence for forest species was + 408m (ranging from -11m to +1900m) and differed significantly from zero (Wilcox test: $p\text{-value} < 0.001$) (Fig. 4). For non-forest species, the mean depth of edge influence was -117m (ranging from -770m to +361m) and also differed significantly from zero (Wilcox test: $p\text{-value} = 0.014$). Estimates of the depth of edge influence for forest and non-forest species were significantly different (Wilcox test: $p\text{-value} < 0.001$). Depth of edge influences did not differ significantly between forest-dependent amphibians and reptiles (mean: 457m and 370m respectively, Wilcox test: $p\text{-value} = 0.5$).

Some non-forest species had a depth of edge influence greater than zero (response type 1, Fig. 2), indicating that these species were also present along forest edges (5 amphibians and 6

259 reptiles, 45% of non-forest amphibian and 22% of non-forest reptile species). Among forest-
260 species, no amphibian and only two reptiles (6% of forest reptile species) ever occurred in the
261 matrix.

262

263 **3.2 Same species, multiple landscapes**

264 Seventeen species were present in multiple landscapes. Of which, 11 showed a similar edge
265 response curves across different forested landscapes. These species could therefore be consistently
266 classified as forest or non-forest species (Fig. 5, species for which response types are all in brown
267 shades or blue shades, i.e. 11 last species from *Oophaga pumillio* to *Anolis sericeus*).

268 However, six species showed different types of abundance response curves to the forest
269 edge in the different landscapes. Two of these species, *Holcosus festivus* (Central American
270 whiptail, a medium-sized ground-dwelling lizard found most commonly in humid and moist
271 lowland forests in Central and South America) and *Thecadactylus rapicauda* (radish-tail gecko, a
272 medium-sized Neotropical gecko with a wide geographic distribution over Central and South
273 America), showed a forest-dependent edge response in one or more landscapes, but had no edge
274 response (i.e. response 0) in another one. Four other species were classified as forest species in
275 some landscapes but as non-forest species in others. *Rhinella marina* (cane toad, also known as the
276 giant Neotropical toad), is a species found in all major IUCN habitat types and listed as invasive in
277 large parts of the world. This species showed three different types of edge response curves in three
278 very different fragmented landscapes (a secondary forest, a cocoa plantation and a *Eucalyptus*
279 plantation). Similarly, *Gonatodes albogularis* showed the same kind of edge response (increasing
280 abundance in the core forest area) in three cocoa plantation landscapes, and a different response
281 (more abundant into the matrix) in a mixed landscape mosaic. Finally, *Anolis lemurinus* (ghost
282 anole, partly arboreal species of Central and South America) showed four different response types
283 in four landscapes, all of which cocoa plantations. *Sphenomorphus cherriei* showed the same edge
284 response in secondary regrowth and two cocoa plantation landscapes, but another edge response

285 curve in the third cocoa plantation. For these species, differences among landscape type cannot
286 explain the inconsistency in edge response.

287

288 **4 Discussion**

289 Our findings provide strong support for the impacts of fragmentation, and in particular of edge
290 effects, on amphibians and reptiles in Neotropical landscapes. By clustering species in their
291 response to forest edges, we can show that over 90% of amphibians and reptile species respond to
292 the forest edge (Fig. 3). We further showed that the abundance of forest species does not increase in
293 forest areas in the immediate vicinity of the edge, but that the response occurs gradually over a
294 spatial gradient that can extend to almost +2000m into the forest (Fig. 4). We suggest that this edge
295 effect is likely to reduce the core area of favourable habitat for forest species, with consequences
296 for estimates of local extinction rates following habitat loss using models based on species-area
297 relationships (Wearn et al., 2012). Even non-forest species were impacted by the presence of forest
298 edges, with 50% of all non-forest species showing an edge effect extending up to 50m into the
299 matrix.

300 Amphibians are the most threatened group in Red List assessments (Hoffmann et al.,
301 2010). Here, we show that there are clear abundance variations of amphibians and reptiles at local
302 scales, that signal causal impacts of ecosystem degradation through forest fragmentation. This, in
303 combination with dispersal and resource limitations, both typical for amphibians and reptiles,
304 should make them useful indicators to monitor the health of forest ecosystems and to evaluate
305 habitat restoration efforts (Carignan and Villard, 2002). Abundance signals of forest-core dependent
306 species for example, could be used as early warning systems by managers to monitor the ecological
307 integrity of forests at the landscape scale.

308 A major meta-analysis using 112 published studies concluded that there was “no strong
309 support for the importance of edge effects for either amphibians or reptiles, with a number of
310 studies finding either no effect, a weak effect, or a species-specific effect with no overall change in

richness” (Gardner et al., 2007a). Our findings, utilising 104 species in the Neotropics, contradicts this conclusion: the forest edges affect the abundance of both amphibians and reptiles, for both forest and non-forest species. We offer three possible explanations for this apparent contradiction. First, only 48% of the studies reviewed by Gardner et al. (2007a) used species abundances as a parameter for measuring edge effects. Instead, most of their studies focussed on species richness, a metric that does not capture fragmentation impacts on population viability and that fails to account for edge-related turnover in species composition (Banks-Leite et al., 2012). Second, we returned to the raw data from original studies and used exactly the same primary statistics to examine species abundance responses to edge effects, a more sensitive approach compared to relying on reported summary statistics derived using different statistical tests as is typical for meta-analytic reviews. Third, there was a notable difference in localities, with just 27% of the studies analysed by Gardner et al. (2007a) located in tropical landscapes, where our datasets come from. It is plausible that tropical amphibians are inherently more sensitive to forest edges than those in temperate landscapes. This could be linked to tropical species being less thermo-tolerant, as they are more likely to experience less temperature variation (Deutsch et al., 2008). Furthermore, thermal performance curves of ectotherms are typically skewed, dropping sharply as temperature rises above the optimum value. Hence, the vast majority of ectotherms would experience heat stress at temperatures above 40°C, a temperature they are likely to exceed in more sun-exposed environments (Kearney et al., 2009) such as forest edges in the tropics. However, our data do not allow us to test for this mechanistic link and it is possible that the timing of the fragmentation process differs in shaping patterns of edge responses observed at a given time point (Metzger et al., 2009).

4.1 Possible drivers of response curves

Species responses to forest edges were likely driven by changes in habitat structure and associated changes in micro-habitats and micro-climates. We based our analyses on forest fragments

delineated from high spatial resolution tree cover maps, which ultimately can be related to canopy structure and, in particular, canopy openness. Dense canopies filter out up to 95% of the incoming solar radiation (Bonan, 2008), limiting light availability on the forest floor and affecting microclimates within the forest (Ashcroft and Gollan, 2012). Air beneath dense forest canopies is cooler and holds higher relative humidity during the day, and forest microclimate is also less variable within dense forests (Hardwick et al., 2015). The shape of the response curve to the forest edge is likely to also depend on the sensitivity of the species or taxonomic group to this variation in micro-habitats, rendering more specialized species more vulnerable to forest fragmentation (McKinney, 1997; Henle et al., 2004). While habitat generalists may show adaptive switching behaviour in response to variation in habitat availability and quality (Wilson and Yoshimura, 1994), habitat specialists, especially when also characterised by reduced thermo-tolerance and low dispersal capacity, may be less able to cope with the heterogeneous canopies and microclimates characterising forest edges or matrix habitats (Didham and Lawton, 1999).

We found that two thirds of the species present in several landscapes responded in a similar fashion to forest edges in all landscapes (Fig. 5). The remaining species, which displayed different response types in different fragmented landscapes, are all habitat generalists. For instance, the cane toad *Rhinella marina* is an “extreme generalist” species (IUCN Red List), occurring in almost all habitat types, and the ghost anole *Anolis lemurinus* was described as “extremely variable in all morphological characters as well as in pattern” (Stuart, 1955). We suggest that habitat generalists are more likely to have inconsistent edge responses among landscapes than specialists because they are better able to exploit non-forest habitats within fragmented landscapes (Henle et al., 2004). For example, the cane toad is present in forest fragments, so could conceivably be classified as a forest species in some locations, but is more abundant in matrix habitats such as sugar cane plantations where it would be classified as a non-forest species. In contrast, specialist species may be more sensitive to forest fragmentation because they are unable to cross major gaps between forest patches in the absence of forest corridors (Hobbs, 1992; Henle et al., 2004).

Although, edge effects show complex interactions with other landscape elements and disturbance regimes. For example, Isaacs-Cubides and Urbina-Cardona (2011) found that the anthropogenic disturbance rather than the edge effects affected the presence of some rare anuran species in a Neotropical mountain cloud forest, affecting the habitat quality and population persistence. Moreover, the edge effect may vary with climatic season, some species appearing as edge-avoiders during wet season and edge-loving species during dry season (Schlaepfer and Gavin, 2001).

The heterogeneity in the distribution of different matrix habitat types among some landscapes may also explain the inconsistency of edge responses of a species across several datasets, as matrix habitats may present different levels of suitability (e.g., a secondary forest might be less unsuitable than a pasture for a forest species). Moreover, the high habitat heterogeneity at a finer scale may affect the edge response curves of some species, especially those from amphibians that are sensitive to the proximity of streams or ponds.

4.2 Implications for biodiversity sampling

Our findings suggest that detecting herpetofaunal responses to forest fragmentation requires assessment of abundance trends, collected over a gradient of distances to forest edge that may stretch for at least 250m into the forest interior and 50m into the matrix. Matrix type and context should be taken into account as they could affect species composition and abundance (Santos-Barrera and Urbina-Cardona, 2011). Habitat quality along forest interior could be affected by the intrusion of matrix species that uses canopy openings as stepping stones, altering interspecific interactions into the forest interior (Sartorius et al. 1999, Urbina-Cardona et al. 2006, Carvajal-Cogollo and Urbina-Cardona 2015).

We found that more than 50% of all forest species were affected by forest edges for at least +250m within the forest interior, suggesting that short edge transects may fail to detect important ecological responses. For example, the Turnip-tailed Gecko *Thecadactylus rapicauda* showed no

edge response in the Montserrat landscape, sampled up to 300m within the forest (Young and Ogradowczyk, 2008), but displayed a clear forest-dependent edge response curve in the Brazilian landscape sampled up to 3780m (Gardner et al., 2007b). However, a rapid assessment of studies on amphibian and reptile edge responses shows that sampling designs rarely include plots farther than a few hundred meters in the forest (e.g. Biek et al., 2002: 75m; Lehtinen et al., 2003: 50m; Reino et al., 2009: 300m; Toral et al., 2002: 250m). This use of relatively short edge transects may have contributed to previous failings to detecting fragmentation impacts on amphibians and reptiles. Ries and Sisk (2010) suggest also that weak research design could lead to “neutral” edge responses (i.e. response type 0).

Similarly, edge transects need to encompass both sides of the habitat edge (Ewers and Didham, 2006a). We found that 50% of non-forest species were affected by the forest edge to at least -50m within the matrix. However, with the exception of one study measuring edge responses (Urbina-Cardona et al., 2012; dataset 1 in Table 1), the sampling design in other studies either ignored the matrix altogether or consistently under-sampled the matrix. For example, Medina-Rangel (2011) compared biodiversity patterns across five habitats (dataset 2, table 1), and hence included some plots in the matrix. However, in the other landscapes, only 12.5% of plots across all studies were placed outside the forest.

We therefore suggest that plots should be sampled from -300m from the edge in the matrix to +1000m from the edge into the forest (if possible given the size of the forest remnants) with particular focus on intense sampling in the zone next to the edge. Information on both matrix and forest structure (their “quality”) should be recorded alongside measures of species abundances. Our findings are also relevant for research carried out in fragments that due to their small size may not allow for sampling to be implemented in this design. We suggest that such small forest fragments are likely to be devoid of forest core habitat but rather represent one extended forest edge habitat, determined by biotic and abiotic variables such as wind, weed invasion, tree mortality or phenology (Broadbent et al., 2008). This edge habitat is useful as a stepping stone or habitat

415 corridor but is unlikely to support forest specialists on the long-term. Studying such small
416 fragments should hence be seen as important for understanding biodiversity response to
417 fragmentation at landscape level, indicating interconnectedness between patches (Ribeiro et al.,
418 2009), but is unlikely to provide mechanistic insight into fragmentation responses of species at
419 patch level.

420 Here, we analysed the influence of a single forest edge, the one nearest each plot. This
421 represents an important limitation in many fragmentation studies, as it is clear that real landscapes
422 encompass a complex mosaic of forest patches separated by various matrix types. This landscape
423 mosaic configuration could partially explain the high diversity of edge response type of some
424 lizards such as *Anolis lemurinus*, *Gonatodes albogularis* or *Sphenomorphus cherriei* (Fig. 5). The
425 shape of the response curve is also likely to be affected by landscape-scale fragmentation
426 characteristics including matrix contrast, patch connectivity and the presence of multiple edges
427 (Ries et al., 2004; Fletcher, 2005). For example, the Mexican landscape in our study consists of
428 forest fragments separated by pasture, facilitating a clear delineation of fragments based on strong
429 contrasts in tree cover. The matrix in the Colombian landscape, on the other hand, includes various
430 non-forest habitat types with a gradient of tree cover contrasts, complicating the delineation of
431 fragments juxtaposed to the surrounding matrix. Modelling abundance changes of species in
432 response to fragmentation based on species' perception of the continuous variation in tree cover
433 across the landscapes would be an interesting direction for future studies.

434 Furthermore, efforts in the field should include measures of biophysical structure, and in
435 particular canopy structure. Rapid canopy structure assessments such as those based on
436 hemispherical photography are increasingly implemented following standardised sampling designs.
437 They allow for rapid and cost-effective retrieval canopy lead area index and fraction of absorbed
438 photosynthetically active radiation (Pfeifer et al., 2012, 2014), which mechanistically link to
439 microclimates (Hardwick et al., 2015). These field estimates can then be up-scaled to landscape
440 structure maps using increasingly available high spatial-resolution satellite data (Pfeifer et al.,

2016), which can be utilised for detailed analyses and interpretation of biodiversity changes induced by fragmentation. Future analyses could use vegetation productivity or tree cover maps created directly from Landsat images acquired as close as possible to the sampling date in the field. Future campaigns sampling biodiversity in fragmented forested landscapes could make use of recent developments in remote sensing and ecological sciences, acquiring measures of canopy cover following standardised sampling designs in the field and using remotely sensed data to up-scale those measures to landscape maps (Pfeifer et al., 2016).

4.3 Conservation implications

In this study, the depth of edge influence was $\geq 250\text{m}$ for at least 50% of the species (Fig. 4). Thus, the edge zone extends on average from 0 to 250m within the forest habitat, so that a round forest patch with a mean diameter $< 500\text{m}$ would contain no viable core area for the forest species. This has strong consequences in terms of conservation, as it clearly shows that large forest patches must be conserved to protect forest species. Otherwise, only species able to use the immediate vicinity of forest edges (i.e. less prone to edge effect) are likely to survive. This is of a particular concern under current road expansion efforts (Laurance et al., 2014), which increasingly cut into fragments the remaining wilderness areas. In the Brazilian Atlantic forest for example, more than 80% of the fragments are $< 50\text{ha}$ and half of the remaining forest is closer than 100m from an edge (Ribeiro et al., 2009), whereas up to 50% of the dry tropical forest in Central and South America remain as fragments $< 10\text{km}^2$ (Portillo-Quintero and Sanchez-Azofeifa, 2010). In our study, the range of depth of edge influence from 200m to 300m into the forest includes almost 60% of the forest species, with 85% of forest species showing a depth of edge influence greater than 200m.

Our approach of computing species response curves to the nearest forest edge explicitly quantifies the depth of edge effect, which ultimately is a measure that can be used to delineate the minimum forest area required to protect forest-dependent and edge-sensitive species in any given landscape. This measure can also be used in ecological networks, i.e. large-scale corridors that

467 connect habitat patches for animal dispersal (Samways et al., 2010): these networks create new
468 edges and it is relevant to know the depth of edge effect on the targeted species in order to design
469 them (Pryke and Samways, 2012). For example, Dixo and Metzger (2009) showed that too narrow
470 corridors may prevent leaf-litter lizards from recolonizing forest fragments.

471 Furthermore, our approach can be used to assess responses of species within a taxonomic
472 group measured in the same landscape. It allows quantifying the forest area needed to maintain the
473 majority of species. This measure can then be used to identify and delineate those areas in land
474 cover maps that meet (or could potentially meet under restoration efforts) the criterion of maximum
475 patch size, and therefore the minimum critical forest area required to maintain forest biodiversity
476 over time. This method is thus particularly relevant in prioritizing landscapes for conservation
477 when several species are involved (Moilanen et al., 2005). Here, 80% of all forest species showed a
478 depth of edge influence narrower than 450m, indicating that a forest patch with a diameter of
479 approximatively 1km (~ 80ha area) provides a suitable core area of at least 3ha (100m diameter)
480 for most edge-affected species.

481 More recently, various indicators have been developed to assess the population abundance
482 dimension of biodiversity, including the Living Planet Index (LPI), the Wild Bird Index (WBI) and
483 the European Butterfly Indicator for Grassland Species (Pereira et al., 2012). Here we show that
484 abundance adds significant information, allowing to develop a more mechanistic understanding of
485 how species respond to land use change and in particular deforestation. This understanding, in turn,
486 will improve our capacity to predict biodiversity change following habitat loss and fragmentation,
487 paving the way for managing biodiversity change in human-modified landscapes. Many studies
488 aiming to predict biodiversity loss following habitat loss and fragmentation continue to link species
489 distributional data to models of species-area relationships. Matthews et al. (2014) showed that
490 focussing on total species richness instead of separating specialists and generalists may
491 underestimate the loss of specialists following fragmentation, whilst overestimating the value of
492 smaller fragments. Here, we go one step further showing that the sensitivity of specialists varies

493 and that abundance signals in response to forest edge can be used to delineate the forest habitat that
494 remains useful for forest specialists. Future studies should include comparisons between both
495 approaches, using species richness and abundance measures, to identify under which situations
496 species richness would be insufficient to plan for management of biodiversity loss at landscape
497 scale.

498

499 **5 Conclusion**

500 Here, we present for the first time strong support for pervasive forest fragmentation impacts on a
501 vast number of amphibian and reptile species in Neotropical forest landscapes. The extent of this
502 edge effect stretches farther than detected by most studies, both within and outside forest remnants,
503 necessitating a rethink of how we sample biodiversity in fragmented landscapes. The edge
504 responses of amphibians and reptiles are likely to depend at least in part on their life-history traits,
505 such as body size and degree of habitat specialization, although this will require considerable
506 further work to demonstrate general patterns of trait-determined edge sensitivity. This may be a
507 promising avenue for future work, however, as it could allow us to identify the species most at risk
508 from future land use changes involving deforestation and forest fragmentation, and ultimately
509 design landscapes that can maximise biodiversity and functional diversity values under land use
510 changes.

511

512 **Acknowledgements**

513 LSM's Master thesis work was funded by the French Ministry for Higher Education and Research
514 and the École Normale Supérieure de Paris. MP, VL and RME were supported by European
515 Research Council Project number 281986. This paper is a contribution to the global BIOFRAG
516 project.

517

518 **References**

- 519 Andren, H., 1994. Effects of habitat fragmentation on birds and mammals of suitable habitat: a
520 review landscapes with different proportions. *Oikos* 71, 355–366.
- 521 Araújo, M.B., Pearson, R.G., 2005. Equilibrium of species' distribution with climate. *Ecography*
522 (Cop.). 28, 693–695.
- 523 Arnaud-Haond, S., Duarte, C.M., Alberto, F., Serrao, E.A., 2007. Standardizing methods to address
524 clonality in population studies. *Mol. Ecol.* 16, 5115–5139.
- 525 Ashcroft, M.B., Gollan, J.R., 2012. Fine-resolution (25 m) topoclimatic grids of near-surface (5
526 cm) extreme temperatures and humidities across various habitats in a large (200 × 300 km)
527 and diverse region. *Int. J. Climatol.* 32, 2134–2148.
- 528 Banks-Leite, C., Ewers, R.M., Metzger, J.P., 2012. Unraveling the drivers of community
529 dissimilarity and species extinction in fragmented landscapes. *Ecology* 93, 2560–2569.
- 530 Bellard, C., Bertelsmeier, C., Leadley, P., Thuiller, W., Courchamp, F., 2012. Impacts of climate
531 change on the future of biodiversity. *Ecol. Lett.* 15, 365–77.
- 532 Biek, R., Mills, L.S., Bury, R.B., 2002. Terrestrial and stream amphibians across clearcut-forest
533 interfaces in the Siskiyou Mountains, Oregon. *Northwest Sci.* 76, 1689–1699.
- 534 Blaustein, A.R., Bancroft, B.A., 2007. Amphibian population declines: evolutionary considerations.
535 *Bioscience* 57, 437.
- 536 Bonan, G.B., 2008. *Ecological Climatology: Concepts and Applications*, Second Edition.
537 Cambridge University Press.
- 538 Broadbent, E.N., Asner, G.P., Keller, M., Knapp, A.E., Oliveira, P.J.C., Silva, J.N., 2008. Forest
539 fragmentation and edge effects from deforestation and selective logging in the Brazilian
540 Amazon. *Biol. Conserv.* 141, 1745–1757.
- 541 Cadenasso, M.L., Traynor, M.M., Pickett, S.T.A., 1997. Functional location of forest edges:
542 gradients of multiple physical factors. *Can. J. For. Res.* 27, 774–782. doi:10.1139/x97-013
- 543 Carignan, V., Villard, M.-A., 2002. Selecting Indicator Species to Monitor Ecological Integrity: A

544 Review. *Environ. Monit. Assess.* 78, 45–61.

545 Carvajal-Cogollo, J.E., Urbina-Cardona, J.N., 2008. Patrones De Diversidad Y Composición De

546 Reptiles En Fragmentos De Bosque Seco Tropical En Córdoba, Colombia. *Trop. Conserv.*

547 *Sci.* 1, 397–416.

548 Carvajal-Cogollo, J.E., Urbina-Cardona, J.N., 2015. Ecological grouping and edge effects in

549 tropical dry forest: reptile-microenvironment relationships. *Biodivers. Conserv.* 24, 1109–

550 1130.

551 Crooks, K.R., 2002. Relative sensitivities of mammalian carnivores to habitat fragmentation.

552 *Conserv. Biol.* 16, 488–502.

553 Cushman, S.A., 2006. Effects of habitat loss and fragmentation on amphibians: a review and

554 prospectus. *Biol. Conserv.* 128, 231–240.

555 Demaynadier, P.G., Hunter, M.L., 1999. Effects of sylvicultural edges on the distribution and

556 abundance of amphibians in Maine. *Conserv. Biol.* 12, 340–352.

557 Didham, R.K., 2010. Ecological consequences of habitat fragmentation. *Encycl. Life Sci.*

558 Didham, R.K., Lawton, J.H., 1999. Edge structure determines the magnitude of changes in

559 microclimate and vegetation structure in tropical forest fragments. *Biotropica* 31, 17–30.

560 Dixo, M., Metzger, J.P., 2009. Are corridors, fragment size and forest structure important for the

561 conservation of leaf-litter lizards in a fragmented landscape? *Oryx* 43, 435.

562 Ewers, R.M., Didham, R.K., 2006a. Confounding factors in the detection of species responses to

563 habitat fragmentation. *Biol. Rev. Camb. Philos. Soc.* 81, 117–42.

564 Ewers, R.M., Didham, R.K., 2006b. Continuous response functions for quantifying the strength of

565 edge effects. *J. Appl. Ecol.* 43, 527–536.

566 Fagan, W.F., Cantrell, R.S., Cosner, C., 1999. How habitat edges change species interactions. *Am.*

567 *Nat.* 153, 165–182.

568 Fahrig, L., 2003. Effects of habitat fragmentation on biodiversity. *Annu. Rev. Ecol. Evol. Syst.* 34,

569 487–515.

570 Fletcher, R.J., 2005. Multiple edge effects and their implications in fragmented landscapes. J.
571 Anim. Ecol. 74, 342–352.

572 Gaffney, S.J., 2004. Probabilistic curve-aligned clustering and prediction with regression mixture
573 models. University of California.

574 Gardner, T.A., Barlow, J., Peres, C.A., 2007a. Paradox, presumption and pitfalls in conservation
575 biology: The importance of habitat change for amphibians and reptiles. Biol. Conserv. 138,
576 166–179.

577 Gardner, T.A., Ribeiro-Junior, M.A., Barlow, J., Avila-Pires, T.C.S., Hoogmoed, M.S., Peres, C.A.,
578 2007b. The value of primary, secondary, and plantation forests for a Neotropical
579 herpetofauna. Conserv. Biol. 21, 775–787.

580 Gibbons, J.W., Scott, D.E., Ryan, T.J., Buhlmann, K. a., Tuberville, T.D., Metts, B.S., Greene, J.L.,
581 Mills, T., Leiden, Y., Poppy, S., Winne, C.T., 2000. The global decline of reptiles, déjà vu
582 amphibians. Bioscience 50, 653–666.

583 Gibbs, J.P., 1998. Distribution of woodland amphibians along a forest fragmentation gradient.
584 Landsc. Ecol. 13, 263–268.

585 Hansen, M.C., Potapov, P. V, Moore, R., Hancher, M., Turubanova, S. a, Tyukavina, A., Thau, D.,
586 Stehman, S. V, Goetz, S.J., Loveland, T.R., Kommareddy, A., Egorov, A., Chini, L., Justice,
587 C.O., Townshend, J.R.G, 2013. High-resolution global maps of 21st-century forest cover
588 change. Science 342, 850–3.

589 Hardwick, S.R., Toumi, R., Pfeifer, M., Turner, E.C., Nilus, R., Ewers, R.M., 2015. The
590 relationship between leaf area index and microclimate in tropical forest and oil palm
591 plantation: Forest disturbance drives changes in microclimate. Agric. For. Meteorol. 201,
592 187–195.

593 Harper, K.A., MacDonald, S.E., Burton, P.J., Chen, J., Brosofske, K.D., Saunders, S.C.,
594 Euskirchen, E.S., Roberts, D., Jaiteh, M.S., Esseen, P.-A., 2005. Edge influence on forest
595 structure and composition in fragmented landscapes. Conserv. Biol. 19, 768–782.

596 Harris, L., Silva-Lopez, G., 1992. Forest fragmentation and the conservation of biological diversity,
597 Conservation Biology. Springer US, Boston, MA.

598 Henle, K., Davies, K.F., Kleyer, M., Margules, C.R., Settele, J., 2004. Predictors of species
599 sensitivity to fragmentation. *Biodivers. Conserv.* 13, 207–251.

600 Hobbs, R.J., 1992. The role of corridors in conservation: Solution or bandwagon? *Trends Ecol.*
601 *Evol.* 7, 389–92.

602 Hoffmann, M., Hilton-Taylor, C., Angulo, A., Böhm, M., Brooks, T.M., Butchart, S.H.M.,
603 Carpenter, K.E., Chanson, J., Collen, B., Cox, N.A., Darwall, W.R.T., Dulvy, N.K., Harrison,
604 L.R., Katariya, V., Pollock, C.M., Quader, S., Richman, N.I., Rodrigues, A.S.L., Tognelli,
605 M.F., Vié, J.-C., Aguiar, J.M., Allen, D.J., Allen, G.R., Amori, G., Ananjeva, N.B., Andreone,
606 F., Andrew, P., Aquino Ortiz, A.L., Baillie, J.E.M., Baldi, R., Bell, B.D., Biju, S.D., Bird, J.P.,
607 Black-Decima, P., Blanc, J.J., Bolaños, F., Bolivar-G W., Burfield, I.J., Burton, J.A., Capper,
608 D.R., Castro, F., Catullo, G., Cavanagh, R.D., Channing, A., Chao, N.L., Chenery, A.M.,
609 Chiozza, F., Clausnitzer, V., Collar, N.J., Collett, L.C., Collette, B.B., Cortez Fernandez, C.F.,
610 Craig, M.T., Crosby, M.J., Cumberlidge, N., Cuttelod, A., Derocher, A.E., Diesmos, A.C.,
611 Donaldson, J.S., Duckworth, J.W., Dutson, G., Dutta, S.K., Emslie, R.H., Farjon, A., Fowler,
612 S., Freyhof, J., Garshelis, D.L., Gerlach, J., Gower, D.J., Grant, T.D., Hammerson, G.A.,
613 Harris, R.B., Heaney, L.R., Hedges, S.B., Hero, J.-M., Hughes, B., Hussain, S.A., Icochea M,
614 J., Inger, R.F., Ishii, N., Iskandar, D.T., Jenkins, R.K.B., Kaneko, Y., Kottelat, M., Kovacs,
615 K.M., Kuzmin, S.L., La Marca, E., Lamoreux, J.F., Lau, M.W.N., Lavilla, E.O., Leus, K.,
616 Lewison, R.L., Lichtenstein, G., Livingstone, S.R., Lukoschek, V., Mallon, D.P., McGowan,
617 P.J.K., McIvor, A., Moehlman, P.D., Molur, S., Muñoz Alonso, A., Musick, J.A., Nowell, K.,
618 Nussbaum, R.A., Olech, W., Orlov, N.L., Papenfuss, T.J., Parra-Olea, G., Perrin, W.F.,
619 Polidoro, B.A., Pourkazemi, M., Racey, P.A., Ragle, J.S., Ram, M., Rathbun, G., Reynolds,
620 R.P., Rhodin, A.G.J., Richards, S.J., Rodríguez, L.O., Ron, S.R., Rondinini, C., Rylands,
621 A.B., Sadovy de Mitcheson, Y., Sanciangco, J.C., Sanders, K.L., Santos-Barrera, G.,

622 Schipper, J., Self-Sullivan, C., Shi, Y., Shoemaker, A., Short, F.T., Sillero-Zubiri, C., Silvano,
 623 D.L., Smith, K.G., Smith, A.T., Snoeks, J., Stattersfield, A.J., Symes, A.J., Taber, A.B.,
 624 Talukdar, B.K., Temple, H.J., Timmins, R., Tobias, J.A., Tsytulina, K., Tweddle, D., Ubeda,
 625 C., Valenti, S. V, van Dijk, P.P., Veiga, L.M., Veloso, A., Wege, D.C., Wilkinson, M.,
 626 Williamson, E.A., Xie, F., Young, B.E., Akçakaya, H.R., Bennun, L., Blackburn, T.M.,
 627 Boitani, L., Dublin, H.T., da Fonseca, G.A.B., Gascon, C., Lacher, T.E., Mace, G.M., Mainka,
 628 S.A., McNeely, J.A., Mittermeier, R.A., Reid, G.M., Rodriguez, J.P., Rosenberg, A.A.,
 629 Samways, M.J., Smart, J., Stein, B.A., Stuart, S.N., 2010. The impact of conservation on the
 630 status of the world's vertebrates. *Science* 330, 1503–9.

631 Holt, R.D., Keitt, T.H., 2005. Species' borders: a unifying theme in ecology. *Oikos* 108, 3–6.

632 Houlahan, J.E., Findlay, C.S., 2003. The effects of adjacent land use on wetland species richness
 633 and community composition. *Wetlands* 26, 79–96.

634 Isaacs-Cubides, P.J., Urbina-Cardona, J.N., 2011. Anthropogenic disturbance and edge effects on
 635 anuran assemblages inhabiting cloud forest fragments in Colombia. *Nat. Conserv.* 9, 1–8.

636 Kearney, M., Shine, R., Porter, W.P., 2009. The potential for behavioral thermoregulation to buffer
 637 “cold-blooded” animals against climate warming. *Proc. Natl. Acad. Sci. U. S. A.* 106, 3835–
 638 3840.

639 Laurance, W.F., Sayer, J., Cassman, K.G., 2014. Agricultural expansion and its impacts on tropical
 640 nature. *Trends Ecol. Evol.* 29, 107–16.

641 Laurance, W.F., Camargo, J.L.C., Luizão, R.C.C., Laurance, S.G., Pimm, S.L., Bruna, E.M.,
 642 Stouffer, P.C., Bruce Williamson, G., Benítez-Malvido, J., Vasconcelos, H.L., 2011. The fate
 643 of Amazonian forest fragments: a 32-year investigation. *Biol. Conserv.* 144, 56–67.

644 Lehtinen, R.M., Ramanamanhato, J.-B., Raveloarison, Jo.G., 2003. Edge effects and extinction
 645 proneness in a herpetofauna from Madagascar. *Biodivers. Conserv.* 12, 1357–1370.

646 Matthews, T.J., Cottee-Jones, H.E., Whittaker, R.J., 2014. Habitat fragmentation and the species-
 647 area relationship: a focus on total species richness obscures the impact of habitat loss on

648 habitat specialists. *Divers. Distrib.* 20, 1136–1146.

649 Maurer, C.R., Raghavan, V., 2003. A linear time algorithm for computing exact euclidean distance
650 transform s of binary images in arbitrary dimensions. *IEEE Trans. Pattern Anal. Mach. Intell.*
651 25, 265–270.

652 McGarigal, K., Cushman, S.A., 2002. Comparative evaluation of experimental approaches to the
653 study of habitat fragmentation effects. *Ecol. Appl.* 12, 335–345.

654 McKinney, M.L., 1997. Extinction vulnerability and selectivity: combining ecological and
655 paleontological views. *Annu. Rev. Ecol. Syst.* 28, 495–516.

656 Medina-Rangel, G.F., 2011. Diversidad alfa y beta de la comunidad de reptiles en el complejo
657 cenagoso de Zapatosa, Colombia. *Rev. Biol. Trop.* 59, 935–968.

658 Mendenhall, C.D., Frishkoff, L.O., Santos-Barrera, G., Pacheco, J., Mesfun, E., Mendoza Quijano,
659 F., Ehrlich, P.R., Ceballos, G., Daily, G.C., Pringle, R.M., 2014. Countryside biogeography of
660 Neotropical reptiles and amphibians. *Ecology* 95, 856–870.

661 Metzger, J.P., Martensen, A.C., Dixo, M., Bernacci, L.C., Ribeiro, M.C., Teixeira, A.M.G., Pardini,
662 R., 2009. Time-lag in biological responses to landscape changes in a highly dynamic Atlantic
663 forest region. *Biol. Conserv.* 142, 1166–1177.

664 Meyer, F., Beucher, S., 1990. Morphological segmentation. *J. Vis. Commun. Image Represent.* 1,
665 21–46.

666 Murcia, C., 1995. Edge effects in fragmented forests: implications for conservation. *Trends Ecol.*
667 *Evol.* 10, 58–62.

668 Otsu, N., 1979. A threshold selection method from gray-level histograms. *IEEE Trans. Syst. Man.*
669 *Cybern.* 9, 62–66.

670 Pardini, R., 2004. Effects of forest fragmentation on small mammals in an Atlantic Forest
671 landscape. *Biodivers. Conserv.* 13, 2567–2586.

672 Pereira, H.M., Navarro, L.M., Martins, I.S., 2012. Global Biodiversity Change: The Bad, the Good,
673 and the Unknown. *Annu. Rev. Environ. Resour.* 37, 25–50.

674 Pfeifer, M., Kor, L., Nilus, R., Turner, E., Cusack, J., Lysenko, I., Khoo, M., Chey, V.K., Chung,
 675 A.C., Ewers, R.M., 2016. Mapping the structure of Borneo's tropical forests across a
 676 degradation gradient. *Remote Sens. Environ.* 176, 84–97.

677 Pfeifer, M., Gonsamo, A., Disney, M., Pellikka, P., Marchant, R., 2012. Leaf area index for biomes
 678 of the Eastern Arc Mountains: Landsat and SPOT observations along precipitation and
 679 altitude gradients. *Remote Sens. Environ.* 118, 103–115.

680 Pfeifer, M., Lefebvre, V., Gardner, T.A., Arroyo-Rodriguez, V., Baeten, L., Banks-Leite, C.,
 681 Barlow, J., Betts, M.G., Brunet, J., Cerezo, A., Cisneros, L.M., Collard, S., D'Cruze, N., da
 682 Silva Motta, C., Duguay, S., Eggermont, H., Eigenbrod, F., Hadley, A.S., Hanson, T.R.,
 683 Hawes, J.E., Heartsill Scalley, T., Klingbeil, B.T., Kolb, A., Kormann, U., Kumar, S., Lachat,
 684 T., Lakeman Fraser, P., Lantschner, V., Laurance, W.F., Leal, I.R., Lens, L., Marsh, C.J.,
 685 Medina-Rangel, G.F., Melles, S., Mezger, D., Oldekop, J.A., Overal, W.L., Owen, C., Peres,
 686 C.A., Phalan, B., Pidgeon, A.M., Pilia, O., Possingham, H.P., Possingham, M.L., Raheem,
 687 D.C., Ribeiro, D.B., Ribeiro Neto, J.D., Douglas Robinson, W., Robinson, R., Rytwinski, T.,
 688 Scherber, C., Slade, E.M., Somarriba, E., Stouffer, P.C., Struebig, M.J., Tylianakis, J.M.,
 689 Tschardtke, T., Tyre, A.J., Urbina Cardona, J.N., Vasconcelos, H.L., Wearn, O., Wells, K.,
 690 Willig, M.R., Wood, E., Young, R.P., Bradley, A. V., Ewers, R.M., 2014. BIOFRAG - a new
 691 database for analyzing BIODiversity responses to forest FRAGmentation. *Ecol. Evol.* 4,
 692 1524–1537.

693 Pfeifer, M., Lefebvre, V., Gonsamo, A., Pellikka, P., Marchant, R., Denu, D., Platts, P., 2014.
 694 Validating and Linking the GIMMS Leaf Area Index (LAI3g) with Environmental Controls
 695 in Tropical Africa. *Remote Sens.* 6, 1973–1990.

696 Pineda, E., Halffter, G., 2004. Species diversity and habitat fragmentation: frogs in a tropical
 697 montane landscape in Mexico. *Biol. Conserv.* 117, 499–508.

698 Portillo-Quintero, C.A., Sánchez-Azofeifa, G.A., 2010. Extent and conservation of tropical dry
 699 forests in the Americas. *Biol. Conserv.* 143, 144–155.

700 Pryke, J.S., Samways, M.J., 2012. Conservation management of complex natural forest and
701 plantation edge effects. *Landsc. Ecol.* 27, 73–85.

702 Reino, L., Beja, P., Osborne, P.E., Morgado, R., Fabião, A., Rotenberry, J.T., 2009. Distance to
703 edges, edge contrast and landscape fragmentation: Interactions affecting farmland birds
704 around forest plantations. *Biol. Conserv.* 142, 824–838.

705 Ribeiro, M.C., Metzger, J.P., Martensen, A.C., Ponzoni, F.J., Hirota, M.M., 2009. The Brazilian
706 Atlantic Forest: How much is left, and how is the remaining forest distributed? Implications
707 for conservation. *Biol. Conserv.* 142, 1141–1153.

708 Ries, L., Fletcher Jr., R.J., Battin, J., Sisk, T.D., 2004. Ecological responses to habitat edges:
709 mechanisms, models, and variability explained. *Annu. Rev. Ecol. Evol. Syst.* 35, 491–
710 522.

711 Samways, M.J., Bazelet, C.S., Pryke, J.S., 2010. Provision of ecosystem services by large scale
712 corridors and ecological networks. *Biodivers. Conserv.* 19, 2949–2962.

713 Santos-Barrera, G., Urbina-Cardona, J.N., 2011. The role of the matrix-edge dynamics of
714 amphibian conservation in tropical montane fragmented landscapes. *Rev. Mex. Biodivers.* 82,
715 19–27.

716 Sartorius, S.S., Vitt, L.J., Colli, G.R., 1999. Use of naturally and anthropogenically disturbed
717 habitats in Amazonian rainforest by the teiid lizard *Ameiva ameiva*. *Biol. Conserv.* 90, 91–
718 101.

719 Saunders, D.A., Hobbs, R.J., Margules, C.R., 1991. Biological consequences of ecosystem
720 fragmentation: a review. *Conserv. Biol.* 5, 18–32.

721 Schlaepfer, M.A., Gavin, T.A., 2001. Edge effects on lizards and frogs in tropical forest fragments.
722 *Conserv. Biol.* 15, 1079–1090.

723 Stork, N.E., Coddington, J.A., Colwell, R.K., Chazdon, R.L., Dick, C.W., Peres, C.A., Sloan, S.,
724 Willis, K., 2009. Vulnerability and resilience of Tropical forest species to land-use change.
725 *Conserv. Biol.* 23, 1438–1447.

726 Stuart, L.C., 1955. A brief review of the Guatemalan lizards of the genus *Anolis*. Museum Zool.
727 Univ. Michigan.

728 Stuart, S.N., Chanson, J.S., Cox, N.A., Young, B.E., Rodrigues, A.S.L., Fischman, D.L., Waller,
729 R.W., 2004. Status and trends of amphibian declines and extinctions worldwide. *Science* (80-
730). 306, 1783–1786.

731 Suazo-Ortuno, I., Alvarado-Diaz, J., Martinez-Ramos, M., 2008. Effects of conversion of dry
732 tropical forest to agricultural mosaic on herpetofaunal assemblages. *Conserv. Biol.* 22, 362–
733 374.

734 Toral, E.C., Feinsinger, P., Crump, M.L., 2002. Frogs and a cloud-forest edge in Ecuador. *Conserv.*
735 *Biol.* 16, 735–744.

736 Urbina-Cardona, J.N., Olivares-Pérez, M., Reynoso, V.H., 2006. Herpetofauna diversity and
737 microenvironment correlates across a pasture-edge-interior ecotone in tropical rainforest
738 fragments in the Los Tuxtlas Biosphere Reserve of Veracruz, Mexico. *Biol. Conserv.* 132,
739 61–75.

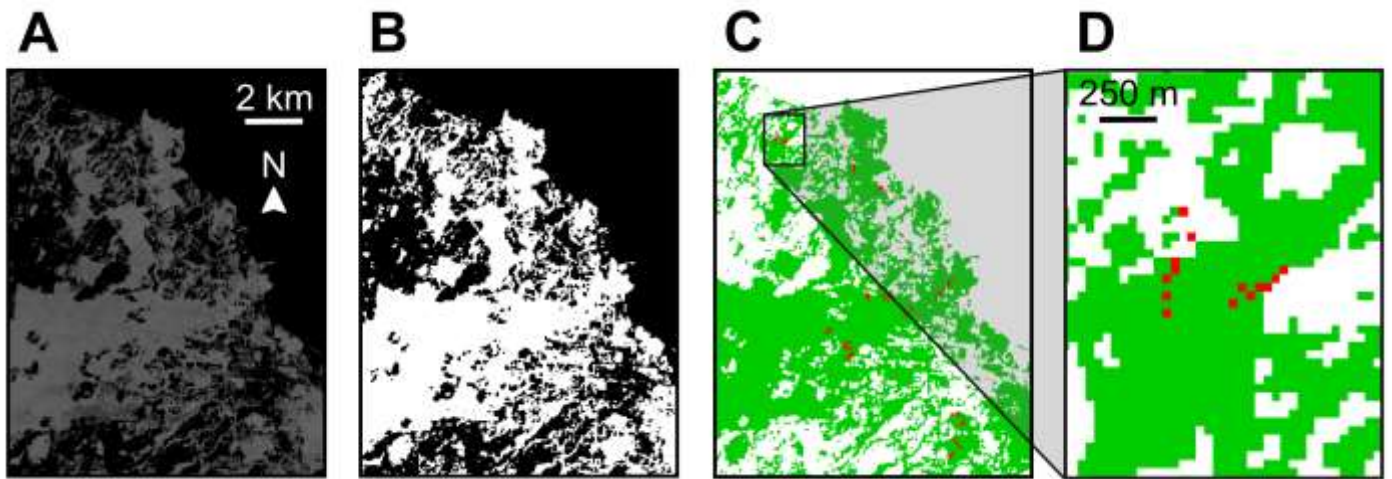
740 Urbina-Cardona, J.N., Reynoso, V.H., Dirzo, R., 2012. Spatio-temporal variation in edge effects on
741 the herpetofauna of a Mexican tropical rain forest, Chapter 3. In: Bezerra, A.D. and Ferreria,
742 T.S. 514 (eds.). *Evergreens: Types, Ecology and Conservation*. Nova Science Publishers.

743 Wearn, O.R., Reuman, D.C., Ewers, R.M., 2012. Extinction debt and windows of conservation
744 opportunity in the Brazilian Amazon. *Science* 337, 228–32.

745 Williams, S.E., Shoo, L.P., Isaac, J.L., Hoffmann, A.A., Langham, G., 2008. Toward an integrated
746 framework for assessing the vulnerability of species to climate change. *Plos Biol.* 6, 2621-
747 2626.

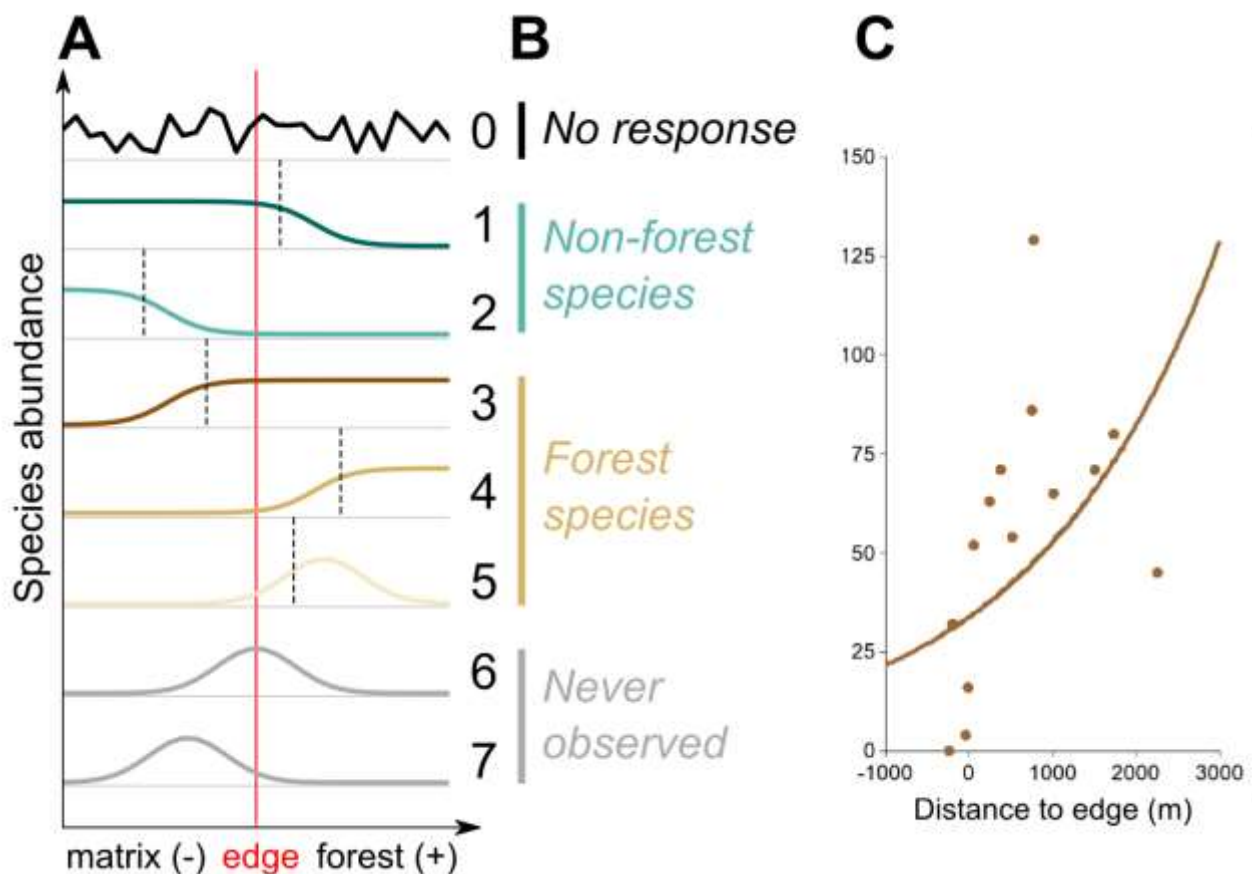
748 Wilson, D.S., Yoshimura, J., 1994. On the coexistence of specialists and generalists. *Am. Nat.* 144,
749 692–707.

750 Young, R.P., Ogrodowczyk, A., 2008. Amphibians and reptiles of the Centre Hills, in: A
751 Biodiversity Assessment of the Centre Hills, Montserrat. p. 90.



752 **Figure 1: Forest fragments delineation and distance to edge computation**

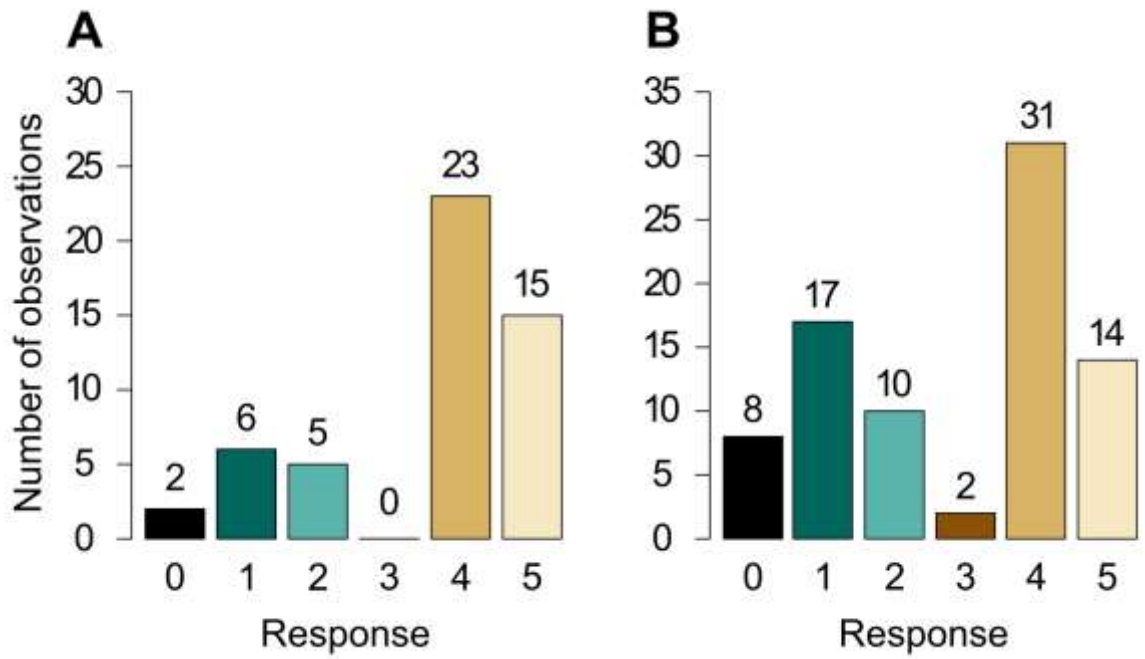
753 The different steps are shown for the dataset #1 in Mexico (cf. Table 1). A: grey scale tree-cover
 754 map from Landsat data (light: high tree cover, dark: low tree cover); B: binary map obtained with a
 755 tree-cover threshold of 41% (forest in white, non-forest in black); C: binary map with plot locations
 756 in red (green: forest, white: non-forest), D: zoom of C to show the distribution of plots across the
 757 forest edge.



758

759 **Figure 2: Different types of edge response (A) and species categories (B)**

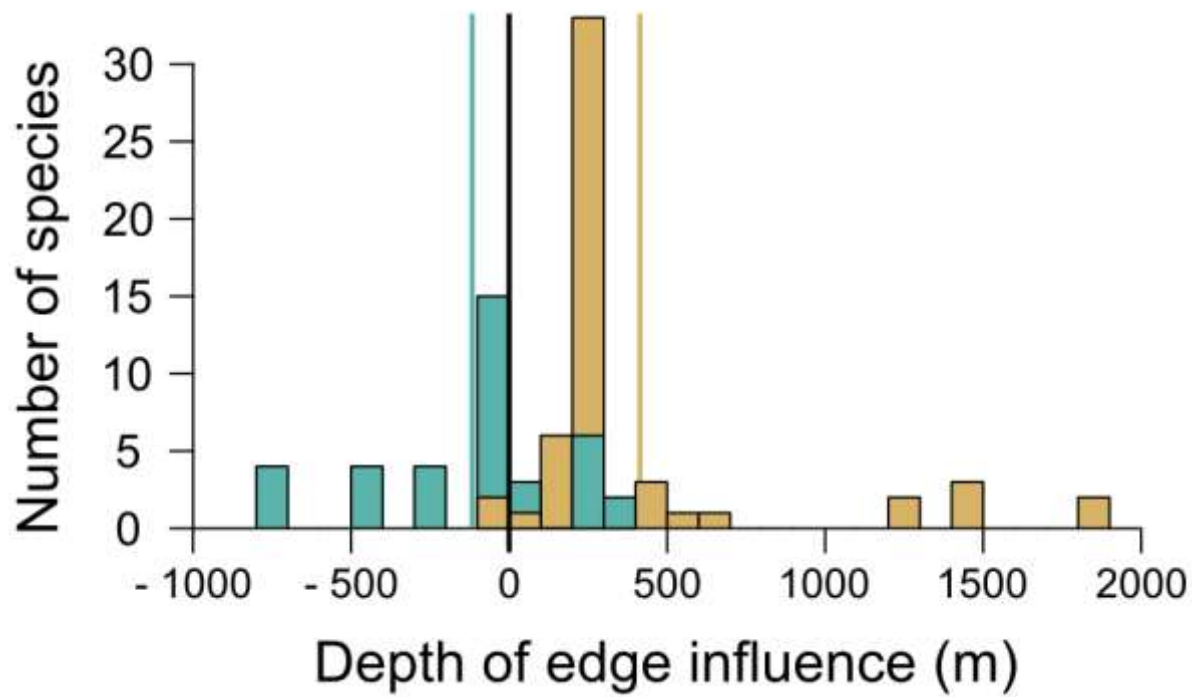
760 There are eight different edge responses, from 0 to 7 (A). Species abundance is directly plotted
 761 with respect to distance to edge. Red line corresponds to the physical edge of the forest; the forest
 762 is on the right of the edge (positive values of distance to edge), the matrix on the left (negative
 763 values) ; the depth of edge effect is shown in dotted line. Species are grouped into categories (B)
 764 regarding to their edge response: 'no response' when edge response is 0 (no clear pattern of change
 765 in abundance across the edge), 'non-forest species' when edge response is 1 or 2 (decreasing
 766 abundance from matrix to forest interior), 'forest species' when edge response is 3, 4 or 5 (increase
 767 in abundance from matrix to forest interior). Edge responses 6 or 7 were never observed. Panel (C)
 768 shows an example of abundance data from dataset #9.



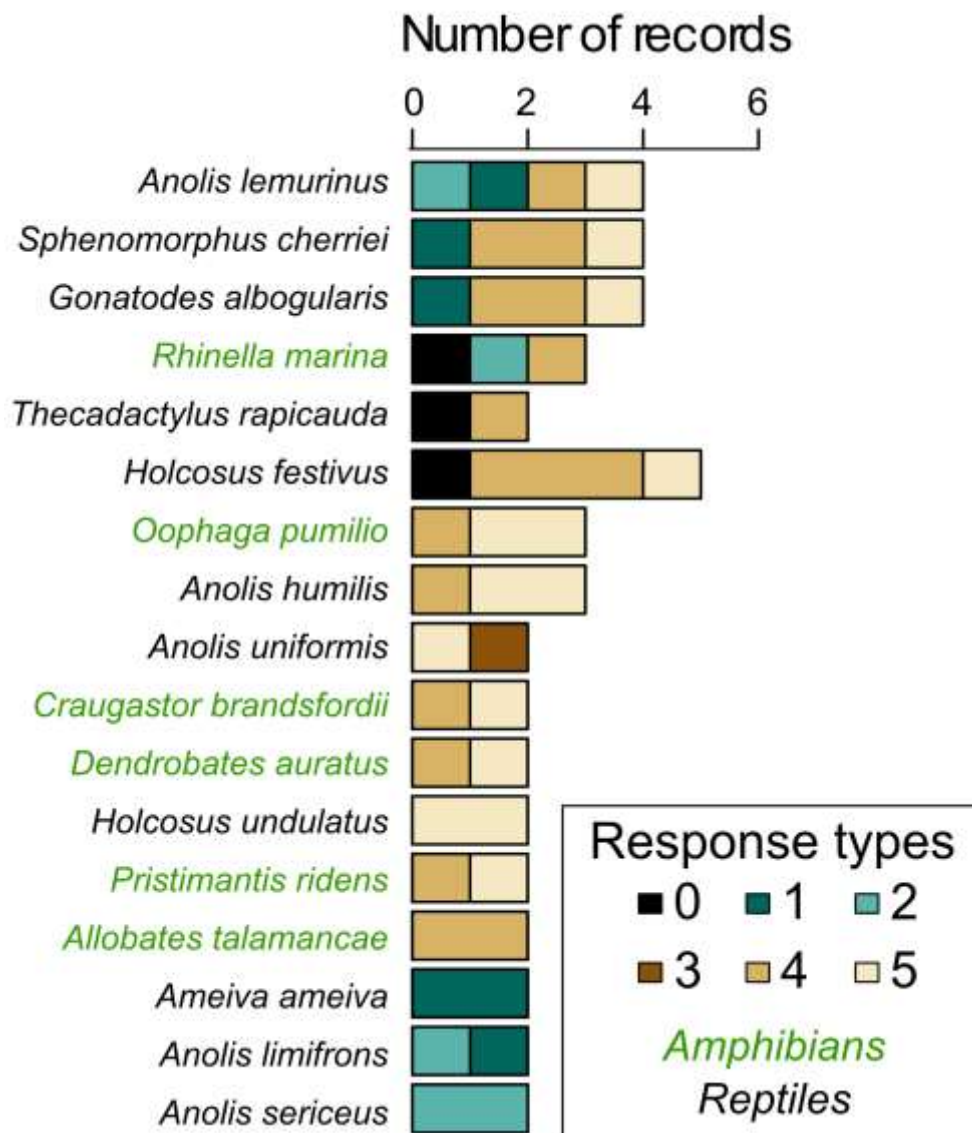
769

770 **Figure 3: Number of recorded edge responses of each type for (A) amphibians and (B)**
771 **reptiles.**

772 Exact numbers of species per edge response type are shown above.



773 **Figure 4: Distribution of the depth of edge influence for forest (brown) and non-forest (blue)**
 774 **species**
 775 Brown line represents the mean depth of edge influence for forest species (+408m), blue line for
 776 non-forest species (-117m) and black line the edge.



777

778 **Figure 5: Edge responses of the 17 species present in several datasets.**

779 Response types indicating a forest-dependence of the species are coloured in shades of brown.

780 Response types indicative of forest independence are coloured in shades of blue. Amphibians

781 names are indicated in green, reptiles names in black.

782 **Table 1: The nine datasets used for analyses**

783 Summary of the datasets characteristics and the mean depths of edge influence on forest-dependent and forest-independent species. ¹Number of
 784 individuals of abundant species in brackets, ²number of abundant species in brackets, ³number of species in brackets.

Data set	Location	Number of plots	Nb. of individuals of known species ¹	Known amphibian species ²	Known reptile species ²	Mean depth of edge influence on forest species (m) ³	Mean depth of edge influence on non forest species (m) ³	Matrix characteristics	Date of sampling	Publication
1	Mexico	162	4332 (4098)	22 (14)	40 (9)	+ 108 (4)	+ 89 (8)	Secondary regrowth or pasture.	2003-2004	Urbina-Cardona et al. 2006, 2012
2	Montserrat	30	3174 (3153)	3 (1)	4 (2)	--	- 85 (2)		2005-2006	Young and Ogradowczyk 2008
3	Colombia	40	795 (713)	0	40 (18)	- 11 (2)	- 163 (10)	Tree-lined savannahs, palm-grove, riparian, dry or chasmophyte forest.	2006-2007	Medina-Rangel 2011
4	Costa Rica	39	725 (658)	19 (6)	23 (8)	+ 441 (7)	- 770 (2)			
5	Guatemala	39	165 (108)	10 (2)	6 (4)	--	--			
6	Panama	43	1251 (1142)	29 (10)	24 (7)	+ 508 (11)	- 244 (4)	Cocoa plantations	2009-2010	Data provided by Eduardo Somarriba
7	Honduras	40	328 (263)	11 (1)	26 (7)	+ 682 (1)	- 247 (6)			
8	Nicaragua	40	458 (422)	12 (6)	18 (7)	+ 167 (3)	- 30 (2)			
9	Brazil	15	2336 (2296)	18 (11)	30 (20)	+ 288 (19)	+ 11 (3)	Primary, secondary and <i>Eucalyptus</i> plantation forests	2004-2005	Gardner et al. 2007b